Numerical Methods to Determine Calcium Release Flux from Calcium Transients in Muscle Cells

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ABSTRACT Several methods are currently in use to estimate the rate of depolarization-induced calcium release in muscle cells from measured calcium transients. One approach first characterizes calcium removal of the cell. This is done by determining parameters of a reaction scheme from a fit to the decay of elevated calcium after the depolarizing stimulus. In a second step, the release rate during depolarization is estimated based on the fitted model. Using simulated calcium transients with known underlying release rates, we tested the fidelity of this analysis in determining the time course of calcium release under different conditions. The analysis reproduced in a satisfactory way the characteristics of the input release rate, even when the assumption that release had ended before the start of the fitting interval was severely violated. Equally good reconstructions of the release rate time course could be obtained when the model used for the analysis differed in structure from the one used for simulating the data. We tested the application of a new strategy (multiple shooting) for fitting parameters in nonlinear differential equation systems. This procedure rendered the analysis less sensitive to ill-chosen initial guesses of the parameters and to noise. A locally adaptive kernel estimator for calculating numerical derivatives allowed good reconstructions of the original release rate time course from noisy calcium transients when other methods failed.

INTRODUCTION

In muscle cells, force development is controlled by Ca²⁺ ions, which are rapidly released from the sarcoplasmic reticulum during sarcolemmal depolarization (Ríos and Pizarro, 1991; Schneider, 1994; Melzer et al., 1995). The calcium concentration change resulting from this release can be measured by using optical indicators, whereas there is no direct way to measure the actual release flux. Various methods have been applied to derive the time course and magnitude of the Ca^{2+} release rate from the measured Ca^{2+} transient (Baylor et al., 1983; Melzer et al., 1984, 1987; Schneider et al., 1985; Brum et al., 1988b; Sipido and Wier, 1991; García and Schneider, 1993; Gonzáles and Ríos, 1993; Pape et al., 1993; Delbono and Meissner, 1996; Shirokova et al., 1996). In these methods, it has to be taken into account that released Ca²⁺ flows to various myoplasmic compartments whose time-dependent Ca²⁺ contents have to be quantified.

Two principally different approaches for estimating calcium release have been described. Both start from kinetic models with binding and transport sites for Ca^{2+} . This leads to a system of nonlinear differential equations. Only the compartment comprised by the indicator is observable. From the indicator signal the concentration of free calcium can be estimated. One approach uses tabulated kinetic parameters of the model and calculates all of the occupancies by using the measured free calcium transient, assuming that

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the literature values apply approximately to the experimental situation (Baylor et al., 1983; Sipido and Wier, 1991; Pape et al., 1993). The other approach tries to determine at least some of the kinetic parameters of the compartment model by analyzing the measured Ca²⁺ signals themselves (Melzer et al., 1984, 1987; Brum et al., 1988b; García and Schneider, 1993; Gonzáles and Ríos, 1993; Delbono and Meissner, 1996; Shirokova et al., 1996). Thereby it is assumed that Ca²⁺ release, which is induced by depolarization, is completely and rapidly stopped after repolarization of the membrane. During the time after repolarization, Ca²⁺ is redistributed among the model compartments. The relaxation time course of free calcium can be predicted with the model equations and compared with the measured time course. This comparison allows us to optimize the parameters in such a way that the calculated relaxation deviates minimally from the measured one. The best fit parameters together with measured Ca2+ transients are then used to calculate the total release as the sum of all Ca²⁺ occupancies at each point in time. The rate of Ca²⁺ release is computed as the time derivative of this sum. Different varieties of this removal fit procedure have been used for analyzing Ca²⁺ measurements from skeletal muscle fibers (Melzer et al., 1986, 1987; Brum et al., 1988a; Gonzáles and Ríos, 1993).

For routine application, this approach requires numerical algorithms that safely lead to a best fit, even when the initial parameter guesses are far from the best ones, and which can handle a certain degree of noise in the experimental data, particularly because derivatives have to be estimated by some smoothing procedure. For this purpose we tested several numerical methods that had been developed for other fields in recent years. For the tests, a simple Ca^{2+} release rate

waveforms were used to simulate free calcium transients similar to those that can be measured in muscle fibers during voltage clamp depolarization (Melzer et al., 1987; Brum et al., 1988b; Feldmeyer et al., 1990). The numerical methods are judged by their ability to reliably determine the input model parameters and release rate time course. We show that the more refined strategies for fitting and smoothing improve the procedure for Ca^{2+} release calculation, and we describe how noise and a violation of the model assumptions can affect the result. Finally, we show that simulation studies can help to decide if a simplified model is still able to yield reliable release rate estimates.

METHODS

Simulation

The model system of ordinary differential equations (ODEs) chosen for this investigation is nonstiff and can be solved numerically by virtually any ODE integrator. For the computations presented here, the routines ODEINT and BSSTEP (Press et al., 1992) were used. Normally distributed random numbers, generated by the routine GASDEV (Press et al., 1992), were added to the simulated transients to produce noisy synthetic data sets.

Parameter estimation

The parameters of the Ca²⁺ dynamics model were estimated from the simulated data by using the method of least squares. The function $f(t, \mathbf{p})$ is the solution at time *t* of the observable part of the model ODE system with the parameter vector \mathbf{p} . Given measurements y_i with error variances σ_i^2 at times t_i , $1 \le i \le N$, the objective function

$$\chi^{2}(\mathbf{p}) = \sum_{i=1}^{N} \left(\frac{y_{i} - f(t_{i}, \mathbf{p})}{\sigma_{i}} \right)^{2}$$
(1)

was minimized with respect to **p**, giving a maximum likelihood estimate for **p**.

Fitting parameters in a system of nonlinear ODEs is cumbersome. As the model function $f(t, \mathbf{p})$ depends nonlinearly on the parameters, the fitting routine has to proceed iteratively, starting from some given initial guess for the parameters. In each iteration, an update step for the estimated parameters has to be computed that will decrease the discrepancy between the model and the given data. The update step is usually computed from the gradient (steepest descent method) or the Hesse matrix (Newton, resp. Gauss-Newton method) of the objective function, or from a combination of both (Levenberg-Marquardt algorithm). The steepest descent method uses a linear approximation of the objective function, whereas the other methods use a quadratic approximation.

In contrast to nonlinear parameter estimation in regression, in ODE systems, no analytical derivatives of the objective function can be supplied. The derivatives can be computed only numerically through finite differences,

$$\frac{\partial f(t_{i}, \mathbf{p})}{\partial p_{j}} \approx \frac{f(t_{i}, p_{1}, \dots, p_{j} + h, \dots, p_{n}) - f(t_{i}, \mathbf{p})}{h}.$$
 (2)

The high computational effort of evaluating $f(t, \mathbf{p})$ usually does not allow for a higher order approximation. Therefore, the choice of *h* can be rather influential. Generally, *h* should be taken as some fractional part, 0.001 say, of p_j , with appropriate modifications if $p_j \approx 0$. The use of internal differentiation can reduce the computational effort substantially (Hairer et al., 1987).

It is well known that ill-chosen initial estimates of the parameters can preclude most routines from yielding any solution at all because the trial trajectory diverges. In an interactive laboratory environment, this poses no principal problems, as the routine can be started again with new initial parameters until convergence is reached. For routine data analysis, however, robust algorithms should be employed.

Because $\chi^2(\mathbf{p})$ shows a nonlinear dependence on the parameters \mathbf{p} , $\chi^2(\mathbf{p})$ will usually have several local minima apart from the true global one that corresponds to the true parameters. Most parameter estimation routines signal convergence of their minimizing iteration if they cannot reduce $\chi^2(\mathbf{p})$ in a small neighborhood of the actual parameter values any further. Thus they might return estimates of \mathbf{p} that are far from the desired true values. If the routine used does not include a strategy for circumventing this problem, every fit has to be inspected by eye to find out whether the desired global minimum has been reached.

Two methods for minimizing $\chi^2(\mathbf{p})$ are compared in this paper.

Initial value approach

If the problem of parameter identification in ODE systems is treated in the same way as other problems of nonlinear regression, one is led to the so-called initial value approach: starting from some initial values for the trial trajectory, the model equations are solved over the whole fitting interval, and $\chi^2(\mathbf{p})$ is calculated. Derivatives are computed by finite differences. Then the parameters are adjusted in such a way as to bring the solution closer to the data.

Quite often, the measurement at the beginning of the fitting interval is taken as a fixed initial value for the solution of the model equations, whereas estimates for the unobserved components are taken from a solution of the ODE system with given parameters along the measurement curve.

For computing the update step, several strategies have been proposed, e.g., the Gauss-Newton method (Schittkowski, 1995) or the Levenberg-Marquardt algorithm (Stortelder, 1996).

Multiple shooting

The technique of multiple shooting has been used for solving boundary value problems in ODE systems (Stoer and Bulirsch, 1993). In the context of parameter estimation, the method was introduced by van Domselaar and Hemker (1975) and, in a much more general context, by Bock (1981, 1983). The observation motivating the use of multiple shooting for parameter estimation is that with the initial value approach, one is effectively neglecting information on the dynamics of the system present in the measurements. Even though the time course of at least one component of the ODE system is known relatively accurately, the initial value approach does not take advantage of any but the very first observation in the fitting interval. If the initial parameters are far from the correct ones, the trial trajectory can deviate arbitrarily far from the measurements.

With Bock's elegant approach, one partitions the fitting interval into many subintervals. The ODE system is solved separately on each of these intervals, using as starting guesses for the initial values the measurements at the beginning of these intervals. None of these initial values are fixed to the observations during the iteration. The initial values at these points have thus become new parameters of the model.

This approach leads to an initially discontinuous trajectory, which is, however, close to the measurements. The final trajectory must of course be continuous, i.e., the computed solution at the end of one subinterval must finally equal the initial value at the beginning of the next subinterval. To enforce this condition, continuity constraints are imposed on the solution. In each iteration, the method has to choose an update step that will not only lead to a minimum of the objective function $\chi^2(\mathbf{p})$, but will also satisfy the continuity constraints in a linearized form. In this way, the newly introduced additional parameters, viz., the initial values at the mesh points, are eliminated from the linearized problem that has to be solved to compute the update step.

Because only the linearized continuity constraints are imposed on the update step, the iteration is allowed to proceed to the final continuous solution through "forbidden ground": the iterates will generally be discontinuous trajectories. This freedom allows the method to stay close to the observed data, prevents divergence of the numerical solution, and reduces the problem of local minima.

As constraints on the parameters or trajectories (e.g., nonnegativity constraints on all rate constants and concentrations in the calcium model) are frequently helpful for further stabilization of the fit, general constraints can be allowed for by taking their linearizations into account when computing the update step. To this end, a method for linear systems with constraints has to be used (Stoer, 1971).

For the reduction of $\chi^2(\mathbf{p})$, a Gauss-Newton method is employed that effectively uses a second-order approximation to $\chi^2(\mathbf{p})$, even though only first derivatives are involved. It can be further stabilized by an appropriate damping strategy that reduces the step size if the problem is highly nonlinear. The rank-deficient case, e.g., when parameters or combinations of parameters cannot be identified reliably, can be treated elegantly by an appropriately chosen rank decision criterion.

Details of the mathematical and implementational aspects of the method are given in Bock (1981, 1983).

Data smoothing and the computation of derivatives of noisy data

To extract the release flux from given Ca^{2+} transients and model parameters, the intracellular Ca^{2+} dynamics has to be reconstructed from the data. This procedure involves integrating the ODE system along the noisy Ca^{2+} measurements and computing the derivative of the measurement curve.

In the present context, the data are interpreted as

$$y_i = f(t_i) + \epsilon_i, \tag{3}$$

where f(t) is the model function and ϵ_i is the measurement error at time t_i , assumed to be normally distributed. When the parameters are known or have been estimated from the data, the smooth curve f(t) underlying the measurements y_i can be identified.

Given data y_i , $1 \le i \le N$, sampled at a resolution of Δt , which are noisy observations of the underlying smooth curve f(t), the derivative $f'(t_i)$ can be estimated by one-sided finite differences as

$$\hat{f}'(t_{\rm i}) = \frac{y_{\rm i+h} - y_{\rm i}}{h\Delta t} \tag{4}$$

or by symmetrical differences as

$$\hat{f}'(t_i) = \frac{y_{i+h} - y_{i-h}}{2h\Delta t},\tag{5}$$

which is equivalent to exploiting the local Taylor series of f(t) to first or second order, respectively. Generalizing this idea, one can try to fit polynomials of a fixed degree k to intervals of the observed data. If the intervals are such that the Taylor series to kth order is a good approximation for f(t), one can take the 0th and first coefficient of the fitted polynomial as an estimate for f(t) and its derivative, respectively. The choice of the correct interval width is essential for the reliability of this method.

With evenly spaced data, such as usually obtained by sampling, this technique can be realized efficiently as a weighted moving average. This method is known as the Savitzky-Golay smoothing filter (Savitzky and Golay, 1964; Press et al., 1992) or as polynomial least-squares convolution (PLSC) (Ratzlaff, 1987). In the context of estimating Ca^{2+} release from Ca^{2+} transients, it has been proposed by Klein et al. (1988).

The Savitzky-Golay smoothing filter is, in fact, a special case of a more general class of methods known as kernel estimators. The basic idea of kernel estimation is to convolve the measurement with a so-called kernel function $K(\theta)$ to obtain estimates $\hat{f}(t_i)$ for the underlying function $f(t_i)$:

$$\hat{f}(t_{i}) = \sum_{j=-b}^{b} y(t_{i+j})K(j).$$
 (6)

Kernel estimators can also be used to estimate derivatives. Kernel functions for derivatives have to satisfy certain moment conditions. Gasser et al. (1985) introduced a family of so-called optimal kernels that is known to have good analytical and practical properties. These kernels were used for the computations in the present paper.

As already noted for the Savitzky-Golay smoothing filter, the choice of the interval that is used to obtain an estimate for f(t) is crucial for the performance of the method. In the context of kernel estimation, the width of this interval, equal to 2b + 1 (cf. Eq. 6), is called the bandwidth.

If b is chosen to be too small, random variations in the data due to observational noise will be "interpreted" as belonging to the signal, thus yielding unsatisfactory results that tend to fluctuate around the true curve. This phenomenon is known as undersmoothing. If, on the other hand, b is chosen to be too large, the curve will be oversmoothed, resulting in the depression of peaks in the data and therefore in a bias of the estimate.

As peaks and even their amplitudes are extremely important in the physiological interpretation of the release rate, oversmoothing can hardly be tolerated. Undersmoothing, on the other hand, can amplify spurious effects in the data to such an extent that their amplitude is of the same size as that of the relevant physiological features.

There are several techniques for choosing the bandwidth b adaptively from the given data (Müller and Stadtmüller, 1987; Seather, 1992). The resulting bandwidth will still lead inevitably to oversmoothing in some parts of the data, whereas other parts will be undersmoothed. One can therefore think of locally adapting b to the structure of the data, decreasing it at peaks and increasing it in flat sections.

Quantitatively, the locally optimal bandwidth of a kernel estimator for the ν th derivative of f(t) is inversely proportional to the square of the (ν + 2)th derivative of f(t), e.g., of the third derivative of f(t) if one wants to estimate f'(t). To take these variations into account, one has to estimate the third derivative of the data. This can be done with an appropriate kernel estimator for which, because of the large variance of the estimate, a rather large bandwidth should be chosen. Mathematical as well as implementational details of this method are given by Müller (1988), whose paper also contains further references to the literature. For the problem discussed here, a fixed bandwidth of 50 ms was chosen to estimate the third derivative.

Boundary effects were treated by generating pseudodata outside the measurement interval as described by Hall and Wehrly (1991), as the behavior of the measured curve long before and long after the depolarization is well known.

Statistics

Confidence intervals for the parameters were obtained from the Hesse matrix,

$$H_{\rm ij} = \frac{\partial^2 \chi^2(\mathbf{p})}{\partial p_{\rm i} \partial p_{\rm j}} \tag{7}$$

of the model equations at the best-fit parameters. It has to be assumed that the model is specified correctly and that the quadratic approximation of $\chi^2(\mathbf{p})$ holds in a large enough region around the solution point. Then the matrix $C = (1/2 H)^{-1}$ is the covariance matrix of the standard errors of the parameters, and the errors of the parameters are multivariately normally distributed with variances c_{ii} (Honerkamp, 1994). Independent 95% confidence intervals for p_i are given by $\hat{p}_i \pm 1.96 \cdot (c_{ii})^{1/2}$.

In some of the simulations presented below we use noiseless data. In these cases, instead of Eq. 1, we minimize the sum over the squared differences between the data and the model predictions. Consequently, no confidence intervals for the parameters can be given.

Hardware and software

The code written for this paper is part of an interdisciplinary project. Because of the different computer platforms involved, portability was a crucial issue. The programs were written in C/C++ and run on IBM RS/6000 workstations, PCs with MS-DOS, and PCs with the LINUX operating system.

Model simulations

For the simulation of artificial Ca^{2+} records, we constructed rate-of-release waveforms, which were used as input flux functions in a simplified Ca^{2+} distribution model.

The calcium release rate in skeletal muscle has been shown to reach a maximum value within a few milliseconds after the onset of a strong depolarization (Baylor et al., 1983; Melzer et al., 1984, 1987; Simon and Schneider, 1988). It then falls to a considerably lower value, probably resulting from a Ca²⁺-dependent inactivation (Schneider and Simon, 1988; Simon et al., 1991; Jong et al., 1993, 1995; Chandler et al., 1995), and finally declines to zero with a very slow time course, because of the combined action of SR depletion (Schneider et al., 1987) and voltage sensor inactivation (Brum et al., 1988b). Even though published release rates vary considerably, the fundamental characteristics, i.e., an early peak followed by a more or less steady level, seem to be conserved in different preparations (Melzer et al., 1987; García and Schneider, 1993; Delbono et al., 1995; Delbono and Meissner, 1996; Shirokova et al., 1996). We constructed release rate records that exhibited a fixed ratio of peak amplitude to a steady level of 2:1 by

using Eq. 8:

$$r(t) = \begin{cases} 0 & \text{for } t \leq 0, \\ s - ae^{-t/\tau_1} + (a - s)e^{-t/\tau_2} & \text{for } 0 < t \leq T, \\ -ae^{-t/\tau_1} + (a - s)e^{-t/\tau_2} + se^{-(t - T)/\tau_r} & \text{for } t > T. \end{cases}$$
(8)

Here τ_1 and τ_2 are the time constants for the rise and decline of the initial peak, respectively. The constant *a* defines an overall scaling of the curve, whereas *s* defines the steady level. The values used for our calculations are given in the legend of Fig. 1.

By varying the depolarization time *T*, release records of different duration were constructed (see Fig. 1 *A*). The decline of the release after repolarization is determined by the time constant $\tau_{\rm r}$, which was set to a low value of 3 ms in all simulations, except the one described in connection with Figs. 9 and 10.

Fig. 1 *B* shows the calcium distribution model that was used for the simulations. It is virtually identical with the one used by Melzer et al. (1986, 1987) to analyze Ca^{2+} removal



FIGURE 1 Construction of synthetic calcium transients. (A) Ca²⁺ release rates calculated with Eq. 8. Parameters: a = 4.6, s = 1, $\tau_1 = 2$ ms, $\tau_2 = 5$ ms. Amplitude 2 μ M/ms. Off time constant, $\tau_r = 3$ ms. *T* had values of 20, 50, 100, 150, and 200 ms to simulate different durations of depolarization. (B) Ca²⁺ distribution model used for the calculations in this paper. (*C*) Synthetic free calcium transients simulated with the model (*B*), Eqs. 9–12, and synthetic release rates (A). Model parameters were as follows: Slow sites: $k_{on} = 10 \ \mu$ M⁻¹s⁻¹; $k_{off} = 0.3 \ s^{-1}$; $S_{total} = 100 \ \mu$ M. Uptake: $k_{ns} = 400 \ s^{-1}$. Expansion due to intrinsic fast sites, F = 30. (*D*) Reuptake of Ca²⁺ into the SR. (*E*) Occupancy of slow sites. (*F*) Occupancy of slow sites on a larger time scale.

and release in voltage-clamped frog muscle fibers, apart from the fact that the Ca^{2+} indicator is not included as a separate compartment in the present model. It was chosen to illustrate the use of the analysis algorithms because it has a small number of parameters and still exhibits the main characteristics of calcium distribution in muscle cells, i.e., fast and slow binding and slow uptake.

The model contains an instantaneously equilibrating nonsaturating compartment CaF, the occupancy of which is a multiple (factor *F*) of the free calcium concentration (Eq. 9), a saturable slow compartment CaS, described by rate constants $k_{\rm on}$ and $k_{\rm off}$ and total concentration of binding sites $S_{\rm total}$ (Eq. 10), and a slow, nonsaturable uptake Ca_{up} with associated fixed rate constant $k_{\rm ns}$ (Eq. 11):

$$C\dot{a}F = F \cdot \dot{C}a_{\text{free}} \tag{9}$$

$$CaS = k_{on} \cdot Ca_{free} \cdot (S_{total} - CaS) - k_{off} \cdot CaS$$
(10)

$$\dot{\mathbf{C}}\mathbf{a}_{\rm up} = k_{\rm ns} \cdot \mathbf{C}\mathbf{a}_{\rm free}.$$
(11)

The time derivatives of the concentrations obey a conservation law:

$$\dot{\mathbf{C}}\mathbf{a}_{\text{rel}}(t) = \dot{\mathbf{C}}\mathbf{a}_{\text{free}}(t) + \mathbf{C}\dot{\mathbf{a}}\mathbf{F}(t) + \mathbf{C}\dot{\mathbf{a}}\mathbf{S}(t) + \dot{\mathbf{C}}\mathbf{a}_{\text{up}}(t), \tag{12}$$

where $\dot{C}a_{rel}(t)$ is the rate of calcium release.

The parameters were chosen to be close to those determined by Melzer et al. (1986) to fit calcium transients measured in frog muscle fibers (see legend of Fig. 1). A peak amplitude of the input calcium release rate of 2 μ M/ms was chosen to generate transients with characteristics similar to those measured in muscle cells (Fig. 1 *C*). A series of five simulations obtained with release durations *T* of 20, 50, 100, 150, and 200 ms was used for the various analyses of this investigation. Fig. 1 shows the release rates (*A*) and the occupancies of the calcium compartments Ca_{free} (*C*), Ca_{up} (*D*), and CaS, the latter one at two different time scales (*E*, *F*).

For the chosen range of release rates, the slow saturable compartment reached saturation levels between 21% and 91%. This leads to a progressive slowing of the relaxation phases in the five free calcium records with increasing release duration. The saturation of this slow compartment reduces its participation in the removal of Ca^{2+} from compartments Ca_{free} and CaF. It is necessary to include in the analysis records with low and with large saturation of CaS, if one wants to precisely identify the parameters k_{on} , k_{off} , and S_{total} .

Determining the release rate

The first step in determining release from given Ca^{2+} transients is a characterization of Ca^{2+} removal. For this purpose, a Ca^{2+} distribution model is used to fit the relaxation phases of measured Ca^{2+} transients. The fit returns a set of kinetic model parameters to describe removal. The only a priori assumption on release is that it is turned off after repolarization. Therefore, it is assumed that the relaxation phases of the free calcium transients reflect redistribution of calcium in and removal from the myoplasmic space involving fluxes between compartments Ca_{free} , CaF, CaS, and Ca_{up} . In general, the model used will at best be a good approximation to the real situation in the cell. In the present case, however, the model structure used for analyzing the simulated calcium transients is identical with the model used for simulation. Therefore, we expect that the values of the four free parameters k_{on} , k_{off} , S_{total} , and k_{ns} can be determined precisely from the information contained in the different calcium relaxation phases (*F* was set to the true value of 30).

Fig. 2 shows that this is indeed the case. Stages 1 and 2 of the figure show the start and end of the iterative process in which a 150-ms interval of each relaxation phase of the five simulated calcium records was fitted with the numerical solution of Eqs. 9–11 with respect to Ca_{free} . Starting values for the fit were as follows: $k_{on} = 5 \ \mu M^{-1} s^{-1}$, $k_{off} = 0.15 s^{-1}$, $S_{total} = 50 \ \mu M$, $k_{ns} = 200 s^{-1}$.

The fitting procedure works in two steps. First, the given calcium transients and initial suggestions for the parameters are used to calculate the occupancies of the nonobservable Ca^{2+} compartments (CaF, CaS, and Ca_{up}). At 15 ms after the start of the release turn-off, the mode of calculation is



FIGURE 2 Removal fit and release calculation. Initial parameters were $k_{\rm on} = 5 \ \mu {\rm M}^{-1} {\rm s}^{-1}$, $k_{\rm off} = 0.15 \ {\rm s}^{-1}$, $S_{\rm total} = 50 \ \mu {\rm M}$, $k_{\rm ns} = 200 \ {\rm s}^{-1}$. All five transients were fitted simultaneously. (1) Thin lines: Synthetic transients. Bold lines: Initial trajectories produced by the fitting routine. (2) Thin lines: Synthetic transients. Bold lines: Trajectories of the fitting routine upon convergence. Best-fit parameters: $k_{\rm on} = 9.95 \ \mu {\rm M}^{-1} {\rm s}^{-1}$, $k_{\rm off} = 0.297 \ {\rm s}^{-1}$, $S_{\rm total} = 100.6 \ \mu {\rm M}$, $k_{\rm ns} = 399.7 \ {\rm s}^{-1}$. True parameters used for simulation: $k_{\rm on} = 10 \ \mu {\rm M}^{-1} {\rm s}^{-1}$, $k_{\rm off} = 0.3 \ {\rm s}^{-1}$, $S_{\rm total} = 100 \ \mu {\rm M}$, $k_{\rm ns} = 400 \ {\rm s}^{-1}$. (3) Occupancy of the model compartments, computed from the transients of (1) using best-fit parameters (shown for the longest depolarization only). (A) Ca_{free} + CaF, (B) CaS, (C) Ca_{up}, (D) Ca_{rel}, equal to the sum of (A), (B), and (C). (4) Release rates computed as the numerical derivative of Ca_{rel} (3D).

changed. Now Eqs. 9–11 are used to calculate free calcium, using the values at the end of the first calculation as initial values for the integration. Because the parameter values are different from the true ones, the calculated curves (*bold lines*) deviate from the simulated calcium records (Fig. 2, stage 1). A least-squares minimization algorithm that searches for a minimum of the objective function $\chi^2(\mathbf{p})$ modifies the parameters until the Euclidean norm of the update step is less than some fractional amount, e.g., 10^{-3} , of the norm of the parameter vector. We do not use relative change of $\chi^2(\mathbf{p})$ as a criterion because numerical experience shows that this may lead to premature signaling of convergence in some cases (Gill et al., 1981).

In the example shown in Fig. 2, it took three iterations for the fitting algorithm to converge (stage 2). As noted in the figure legend, the parameters found by the fit approximated the true parameters with deviations of less than 1%. Convergence to the absolute minimum of the objective function depends on the choice of the parameter starting values. If these starting values are too far from the true values, the algorithms might proceed to a local minimum comprising considerably worse fits or might even diverge. A substantial improvement can be obtained by using the so-called multiple shooting approach introduced by Bock (1981).

Based on the best-fit parameters, the occupancies of all compartments are calculated. Their sum yields the released calcium (Fig. 2, stage 3). Finally, the rate of release is given by the derivative of the released calcium (Fig. 2, stage 4).

RESULTS

In this section we first compare the initial value and the multiple shooting approach for parameter estimation in differential equations. Then we investigate the different smoothing algorithms to estimate derivatives of noisy data. Finally, we examine how violations of the model assumptions influence the estimated rate of release.

Comparison of two fit algorithms

We first compare the two algorithms with respect to their convergence behavior if the starting guesses for the parameters are varied.

Application of the multiple shooting procedure is exemplified in Fig. 3. For this figure, the starting values were chosen to be substantially off the true parameter values: $k_{\rm on} = 1 \ \mu {\rm M}^{-1} {\rm s}^{-1}$, $k_{\rm off} = 1 \ {\rm s}^{-1}$, $S_{\rm total} = 1 \ \mu {\rm M}$, $k_{\rm ns} = 1 \ {\rm s}^{-1}$.

Given these starting guesses for the parameters, the initial value approach (*left*) was unable to return estimates for the parameters, as the trajectory of one of the unobserved compartments in the model diverged during the iteration. The multiple shooting algorithm (*right*), however, succeeded after 11 iterations (Fig. 3 F) and estimated the parameter values with the same precision as in the case of Fig. 2.

As expected, this algorithm proved its superiority when the deviation of the starting values from the true parameter



FIGURE 3 Comparison of initial value approach and multiple shooting approach. Fit of the synthetic transients of Fig. 1, using the ill-chosen initial parameters $k_{on} = 1 \ \mu M^{-1} s^{-1}$, $k_{off} = 1 \ s^{-1}$, $S_{total} = 1 \ \mu M$, $k_{ns} = 1 \ s^{-1}$. All five transients were fitted simultaneously. Thin lines: Synthetic transients. Bold lines: Trajectory produced by the fitting routine. *Left*: Initial value approach. The figure shows iterations 1 (*A*), 2 (*C*), and 3 (*E*). After the third iteration, the program stopped because of the divergence of one of the trajectories of the unobserved compartments. *Right*: Multiple shooting approach with five subintervals. The figure shows iterations 1 (*B*), 3 (*D*), and 11 (*F*). The program signaled convergence after the eleventh iteration. Note that the trajectory is discontinuous during iterations 1 and 3.

values was large. Table 1 summarizes the results of a number of runs with the two algorithms in which different sets of starting values were tried. Furthermore, we compared the performance of the algorithms with respect to positivity constraints on the parameters and investigated the effect of fixing the first point of the trial trajectory to the observation. The entries denote the numbers of iterations until convergence. In several cases, the procedure stopped because the trial trajectories diverged (D). In one instance, no convergence was obtained within 100 iterations (N). A local minimum in parameter space was reached twice (L). When no constraints were imposed on the parameters, both routines failed to converge in a large number of cases. Furthermore, fixing the first fitting point (FIX) to the observation proved to be destabilizing. Thus positivity constraints and a freely varying first point (FREE) should be included in any strategy. As can be seen from the fifth column of Table 1, the multiple shooting approach was able to handle much larger deviations of the starting values from the true parameters than the initial value approach. In the one case in which the multiple shooting algorithm failed, the introduction of small positive (instead of zero) lower bounds on the parameters led to convergence, whereas the initial value approach still diverged.

The multiple shooting algorithm proved to be especially helpful when noisy records were analyzed. Here the fact that the initial values of the trajectory are also optimized was of

TABLE 1	Comparison of initial value and multiple shooti	ng
approache	s for different starting guesses of the parameter	ers

	No constraints		Positivity constr.		
Starting parameters	FIX	FREE	FIX	FREE	Method
	2	2	2	2	IV
10, 0.3, 100, 400	5	5	5	5	MS3
	5	5	5	5	MS5
	3	3	7	7	IV
20, 0.6, 200, 800	6	6	9	8	MS3
	6	6	8	8	MS5
	3	3	8	13	IV
5, 0.15, 50, 200	6	6	5	6	MS3
	6	6	5	5	MS5
	D	D	D	D	IV
0.1, 0.1, 100, 100	D	D	8	13	MS3
	D	D	D	D	MS5
	D	D	12	D	IV
1, 1, 1000, 1000	D	D	8	9	MS3
	D	D	11	11	MS5
	D	D	D	D	IV
1, 1, 100, 100	9	6	8	6	MS3
	9	D	7	7	MS5
	D	D	Ν	L	IV
1, 1, 10, 10	D	D	10	7	MS3
	D	D	9	9	MS5
	D	D	D	L	IV
1, 1, 1, 1	D	D	D	12	MS3
	D	D	D	11	MS5

True values: $k_{on} = 10 \ \mu M^{-1} s^{-1}$, $k_{off} = 0.3 \ s^{-1}$, $S_{total} = 100 \ \mu M$, $k_{ns} = 400 \ s^{-1}$. The parameter *F* was fixed at 30. MS3 and MS5 denote multiple shooting with three and five subintervals, respectively. The table gives the number of iterations needed for convergence. L: Convergence to a local minimum in parameter space; D: divergent behavior; N: no convergence within 100 iterations. Results are shown for either no constraints or positivity constraints on the parameters. Furthermore, the first fitting point was either fixed (FIX) or treated as an additional free parameter (FREE). IV, Initial value; MS, multiple shooting.

considerable advantage. The five artificial Ca²⁺ records of Fig. 1 were subjected to noise by adding normally distributed random numbers with standard deviations σ ranging from 0.005 to 0.2 μ M, and the largest amplitude reached in the calcium transients was 1.5 μ M. Fig. 4 shows examples



FIGURE 4 Synthetic Ca²⁺ transients with different noise levels. Differently scaled identical realizations of normally distributed random numbers were added to the smooth transients shown in Fig. 1 *C*. (*A*) $\sigma = 0.01 \ \mu$ M. (*B*) $\sigma = 0.05 \ \mu$ M. (*C*) $\sigma = 0.1 \ \mu$ M. (*D*) $\sigma = 0.2 \ \mu$ M.

Fig. 5 *A* shows the effect of fixing the first point of the fitted trajectory to the observation at the beginning of the fitting interval. As this value deviates far from the underlying true curve, the final trajectory could not reproduce the data adequately. In the example shown, the release rates computed with the parameters obtained from this fit were unsatisfactory. If the initial value was allowed to vary freely (Fig. 5 *B*), the fit reproduced the data adequately, and the true parameters could be identified within the predicted error margins.

Table 2 summarizes the result of fits to the same original records but subjected to different levels of noise, and compares the results for the two algorithms considered. In each case, three different variants were investigated (for details, see table legend). The multiple shooting algorithm was superior to the conventional initial value approach, in that it produced convergence more reliably at larger noise levels.

Smoothing algorithms and numerical derivatives

The second stage in the rate of release recalculation involves the following step. The model equations, now sup-



FIGURE 5 Removal fit of noisy synthetic calcium transients. All five records of Fig. 4 *D* ($\sigma = 0.2 \ \mu$ M) were fitted simultaneously; only three records are shown. Thin lines, Simulated noisy transient. Dashed lines, Underlying noise-free transient. Bold lines, Best-fit trajectory. (*A*) The initial value of the ODE system was fixed to the measurement at the beginning of the fitting interval. Best-fit parameters (\pm standard errors): $k_{on} = 5 \pm 6 \ \mu$ M⁻¹s⁻¹, $k_{off} = 0 \pm 0.004 \ s^{-1}$, $S_{total} = 266 \pm 447 \ \mu$ M, $k_{ns} = 0 \pm 761 \ s^{-1}$. (*B*) The initial value of the ODE system was allowed to vary freely. Best-fit parameters: $k_{on} = 13 \pm 4 \ \mu$ M⁻¹s⁻¹, $k_{off} = 0.04 \pm 0.18 \ s^{-1}$, $S_{total} = 144 \pm 32 \ \mu$ M, $k_{ns} = 350 \pm 77 \ s^{-1}$.

TABLE 2 Convergence behavior of initial value and multiple shooting approaches at six different noise levels σ

σ (μ M)	IVa	IVb	IVc	MS3	MS5	MS9
A: Paramet	er starting	values: 5,	0.15, 50,	200		
0.005	8	D	9	5	5	6
0.01	8	8	8	5	5	7
0.02	8	10	9	5	5	9
0.05	D	9	L	6	6	7
0.1	D	10	8	8	7	10
0.2	D	D	12	41	11	15
B: Paramet	er starting	values: 20	, 0.6, 200.	800		
0.005	7	8	7	10	9	7
0.01	8	8	7	10	9	7
0.02	9	9	7	10	9	8
0.05	D	D	8	22	8	8
0.1	D	D	Ν	9	7	7
0.2	D	D	L	8	7	8

Two different sets of parameter starting values were chosen for the iteration (A, B). True model parameters: $k_{\rm on} = 10 \ \mu M^{-1} s^{-1}$, $k_{\rm off} = 0.3 \ s^{-1}$, $S_{\rm total} = 100 \ \mu M$, $k_{\rm ns} = 400 \ s^{-1}$, *F* fixed at 30. Starting values for the four free parameters are given in the same sequence with the same units.

IVa, No constraints on the parameters, first point fixed to observed value; IVb: as IVa, but first point as additional free parameter; IVc: as IVb, but with positivity constraints on all parameters.

MS3, MS5, and MS9 denote multiple shooting with positivity constraints on all parameters and with three, five, and nine subintervals, respectively. The table gives the number of iterations needed for convergence. L: Convergence to a local minimum in parameter space; D: divergent behavior; N: no convergence within 100 iterations.

IV, Initial value; MS, multiple shooting.

plied with the best-fit values of the free parameters, are used to calculate the Ca²⁺ occupancies of CaF, Ca_{up}, and CaS by using as the input function a given calcium transient (i.e., the observed function representing Cafree). All four compartment concentrations are summed (Fig. 2, stage 3). Assuming that no Ca^{2+} was present initially, this yields the estimated time course of the total calcium released. To derive the rate of release, a function reflecting the gating of the release channels, the summed total calcium has to be differentiated numerically (Fig. 2, stage 4). This can easily be done by using a difference approximation (Eq. 5, Methods) if the records are smooth. However, as soon as noise is involved, the numerical derivative poses severe problems. To a certain degree this can be overcome by means of signal averaging. Yet, because not all preparations allow for timeconsuming signal averaging, one has to look for optimal procedures to carry out the analysis with noisy records. In the following, we compare the quality of three different ways of estimating numerical derivatives and describe the errors in the calculated rate of release introduced by each one. The first procedure is the conventional difference approximation (Eq. 5). The second method (kernel estimation with globally chosen bandwidth) smoothes the noisy record by a weighted moving average, which is equivalent to fitting polynomials locally. This has the advantage that the derivative can be calculated analytically from the fitted parameters. The choice of the interval length over which each local

fit is carried out results from an analysis of the noise variance σ^2 .

A more refined method (local adaptive algorithm; see Müller, 1988) adjusts the interval length by estimating the local curvature of the signal by means of a crude estimate of the third derivative of the record. This reduces oversmoothing at locations where the signal changes steeply and uses larger smoothing intervals in regions where the curvature is small.

The details of the procedure are demonstrated in Fig. 6. Here, the numerical derivative of a simulated free calcium transient with Gaussian noise ($\sigma = 0.1 \ \mu$ M, comp. Fig. 4 C) is calculated. The original record (with and without noise) is shown in Fig. 6 A. Fig. 6 B shows the estimated third derivative used in the kernel estimator, calculated with a fixed bandwidth of 50 ms (see Methods), and Fig. 6 C shows the resulting kernel bandwidth for each point in time chosen by the algorithm. The continuous line in Fig. 6 Dshows the first time derivative of the noisy calcium transient determined with the locally varying kernel bandwidth of Fig. 6 C. For comparison, the dashed line shows the true derivative of the noise-free transient. The positive peak at the beginning of the time course is underestimated by 23%. This difference results from the trade-off between bias and variance of the estimator. Choosing a smaller bandwidth in this region would decrease the bias but increase the variance. The chosen bandwidth minimizes the expected meansquare error, which combines bias and variance.

Fig. 7 compares a number of methods for calculating the numerical derivative in the determination of the rate of calcium release. Three different noise levels in the simulated calcium records were applied (see legend). To exclude effects due to different realizations of the noise, we used identical realizations scaled to different amplitudes (Fig. 7



FIGURE 6 Computation of the numerical derivative of a noisy free Ca²⁺ transient by locally adaptive kernel estimation. (*A*) Synthetic noisy Ca²⁺ transient ($\sigma = 0.1 \ \mu$ M, cf. Fig. 4 *C*). (*B*) Estimate of third derivative (in μ M/ms³). (*C*) Locally chosen kernel bandwidth (*solid line*). The globally optimal bandwidth (*dashed line*) is 36 ms. (*D*) First derivative of the transient shown in *A*. Solid line: Derivative computed by locally adaptive kernel estimator (in μ M/ms). Dashed line: Analytical derivative of the noise-free transient.



FIGURE 7 Comparison of numerical derivative procedures for the calculation of the rate of calcium release from calcium transients subjected to different levels of noise. The true values of the model parameters were used in each case; therefore, the variability results exclusively from the different ways of calculating the derivatives. (A) Noisy synthetic transients. From left to right: $\sigma = 0.005$, 0.05, and 0.1 μ M. (*B*–*G*) Original (*dashed line*) and recalculated (*continuous line*) rate of release using the following methods of carrying out numerical derivatives of summed total calcium concentration (see Fig. 2): (B) Local adaptive kernel estimator. (*C*) Kernel estimator with optimal global bandwidth (*left*, 8 ms; *middle*, 27 ms; *right*, 36 ms). (*D*) Kernel estimator with fixed bandwidth of 10 ms. (*E*) Kernel estimator with fixed bandwidth of 50 ms. (*F*) Finite differences, h = 4 (see Eq. 5). (*G*) Finite differences, h = 10.

A). In each case, the true values of the model parameter were used for the calculation of the compartment occupancies to exclude variability due to different quality of the removal model fit. Fig. 7 *B* shows the application of the locally adaptive approach outlined in Fig. 6. It produces a rather smooth record and yet gives a relatively good reconstruction of the rapid components of the original rate of release curve (*dashed line*), even for the highest level of noise. From left to right, the calculated peak amplitudes were 98%, 83%, and 86% of the original one. In contrast, a kernel estimator with optimal global bandwidth (Fig. 7 *C*) considerably underestimated the initial peak rate for the larger levels of noise (99%, 67%, 66%). Lowering the global bandwidth in an attempt to get better peak restoration also considerably increased the noise (Fig. 7 *D*). This was

even more the case when a difference approximation of the numerical derivative was used. A choice of the bandwidth (h = 4) that gave good peak restoration completely failed at the higher noise levels (Fig. 7 *F*), and increasing the bandwidth to lower the noise (h = 10) strongly suppressed the peak (Fig. 7 *G*).

Fig. 8 demonstrates the outcome of the complete analysis procedure in the presence of noise. It compares the standard analysis used in the original description of the procedure (Melzer et al., 1987), i.e., the initial value method for parameter estimation by removal fit and finite differences for derivatives on one hand (*left column*), with the multiple shooting method combined with the adaptive kernel estimator on the other hand (*right column*). Apart from the major problems regarding the numerical derivatives, the standard procedure introduced further deviations in the release calculation due to the unreliability in estimating the model parameters and failed to calculate a tolerable estimate of the rate of release time course. The new methods, however, led to a result that stayed close to the original record, even under these unfavorable conditions.

In summary, whereas for relatively clean records (*left column* of Fig. 7) even a difference approximation with small bandwidth (Fig. 7 F) gives good results, the only method that could cope with the higher noise levels was the locally adaptive kernel estimator (Fig. 7 B).

The latter method offers a more reliable estimation of the release rate. This holds for the reconstruction of the peak at the beginning of the depolarization, as well as for the flat parts of the release rate time course.



FIGURE 8 Comparison of two analysis procedures for determining the rate of Ca²⁺ release from the noisy set of free calcium data of Fig. 4 *C* ($\sigma = 0.1 \ \mu$ M). *Left*: Canonically applied method. *Right*: Improved methods. (*A*) Result of removal fit with the initial value approach with no constraints on the parameters and with the first point at the beginning of the fitting interval fixed to the observation. Best fit parameters: $k_{on} = 3.4 \pm 2.0 \ \mu$ M⁻¹s⁻¹, $k_{off} = -0.01 \pm 0.06 \ s^{-1}$, $S_{total} = 342 \pm 289 \ \mu$ M, $k_{ns} = -83 \pm 381 \ s^{-1}$. (*B*) Release rate computed by symmetrical finite differences, h = 10 (Eq. 5). (*C*) Final trajectory of the multiple shooting approach (five subintervals) with positivity constraints on the parameters and with the first point at the beginning of each fitting interval as an additional free parameter. Best fit parameters: $k_{on} = 11 \pm 2 \ \mu$ M⁻¹s⁻¹, $k_{off} = 0.14 \pm 0.11 \ s^{-1}$, $S_{total} = 118 \pm 17 \ \mu$ M, $k_{ns} = 376 \pm 43 \ s^{-1}$. (*D*) Release rate computed by the locally adaptive kernel estimator.

Violation of the model assumptions

A crucial assumption for the removal model fit analysis is that release can be turned off rapidly by repolarization. Only this enables an independent characterization of removal by inspecting the decay of free calcium. If release is still present during the decay, one can expect that removal will be underestimated, and the calculated release time course during the depolarization will be in error. To assess this possible error, we increased the turn-off time constant τ_r of the input calcium release signals (Eq. 8) and analyzed the resulting free calcium transients in the same way as before. Fig. 9 shows the result of this analysis for four different turn-off time constants. Although the peak of the release rate was not affected, increasing τ_r produced an increasingly stronger, slowly decaying undershoot below the actual plateau level. Therefore, delayed closure of the release channels would cause an apparent increase in the ratio of peak to steady level of the determined rate of release.

Fig. 10 explains this result. The figure compares the contribution of the different calcium compartments to the total release rates derived with the removal model fit in two different cases. On the left a rapid ($\tau_r = 3 \text{ ms}$) and on the right a slow ($\tau_r = 25 \text{ ms}$) turn-off of the release had been used for the calcium transient simulation. For the slowly ceasing release, 55% of the steady-state level of release was still active at the time the fit started.

As the model used for fitting assumes that release has turned off completely 15 ms after repolarization, this condition is forced upon the estimated release rate, thereby distorting its shape. The slow saturable component in particular is too small (Fig. 10, *right*).

Inspecting the best-fit parameter values (Fig. 10 legend) shows that this is mainly due to a significant underestimation of S_{total} (43 versus 101 μ M). An additional underestimation of the nonsaturable removal parameter k_{ns} (330



FIGURE 9 Consequences of delayed turn-off of release for the analysis result. Free calcium transients were simulated with release rates with different τ_r (cf. Eq. 8). These transients were then fitted with the removal model, assuming that the release had turned off completely 15 ms after repolarization. Solid lines, release rates computed from the transients using best-fit parameters. Dashed lines, original release rates. The fractions of release still present at 15 ms after repolarization, as a percentage of the steady level (1 μ M/ms), were as follows (τ_r in parentheses): (A) 3% (3 ms); (B) 12% (7 ms); (C) 37% (15 ms); (D) 55% (25 ms).



FIGURE 10 Result of release rate calculation for Ca²⁺ transients simulated with fast and slow turn-off of release. Five calcium transients were generated from synthetic release rates with different τ_r (cf. Eq. 8). These transients were then fitted with the removal model, and release rates were computed with the resulting best-fit parameters. *F* was fixed at 30 in each case. *Left*: $\tau_r = 3$ ms, i.e., fast release turn-off. Best-fit parameters: $k_{on} = 9.95 \ \mu M^{-1} s^{-1}$; $k_{off} = 0.29 \ s^{-1}$; $S_{total} = 100.6 \ \mu M$; $k_{ns} = 399.7 \ s^{-1}$. *Right*: $\tau_r = 25$ ms, i.e., slow release turn-off. Best-fit parameters: $k_{on} = 10.4 \ \mu M^{-1} s^{-1}$; $k_{off} = 0 \ s^{-1}$; $S_{total} = 43.4 \ \mu M$; $k_{ns} = 330 \ s^{-1}$. (A) Simulated transients (*thin lines*) and trajectory from the removal fit (*bold lines*). (B) Total release rates. (C) Rates of change for total fast calcium, i.e., Ca_{free} + CaF. (D) Rates of Ca²⁺ uptake, Ca_{up}. (E) Rates of change for slow calcium, CaS. In *B–E*: Solid lines: Computed. Dashed lines: Original.

versus 400 s⁻¹) contributes little to the underestimation of the removal rate (Fig. 10 *D*, *right*).

Effects of misspecified removal models on the estimated release

In the preceding paragraphs we investigated how the quality of the release rate estimate obtained from the removal model fit approach depends on factors like the level of noise in the data, the numerical procedure used for calculating derivatives, and the degree to which the assumption that release stops immediately after repolarization is fulfilled.

To ensure that the model used for the fit could reproduce the characteristics of Ca^{2+} removal in the system underlying the calculated free calcium data, we used exactly the same model for generating the data. When measured data are analyzed, however, the model used for the fit will at best be an approximation to the real system. In general, the models will be misspecified. This misspecification might show up in different ways:

- 1. The model does not contain as many compartments as the real system.
- 2. Some parameters of the model are fixed to values that are not equal to the true ones.
- 3. The kinetic scheme of the model does not correspond to the real system.

There is no straightforward way to decide if the model used for the fit can sufficiently reproduce the main characteristics of Ca^{2+} release in the experimental system. The criterion generally used to decide if a given model suffices for the analysis is the quality of the removal fit. A model that does not lead to a good fit of the relaxation time course can be assumed to be insufficient for the release calculation. On the other hand, it is assumed, even though not unambiguously proved, that a model which leads to a good fit of the Ca^{2+} relaxation time course after repolarization under different conditions will correctly describe the overall removal properties of the cell and will, therefore, lead to a good approximation of the release rate time course. We believe that this criterion needs further evaluation in future investigations.

It seems more straightforward to decide if a given removal model (which fits the real data well) can be replaced by a simpler model. Here, too, the simulation approach offers a valuable tool. Free calcium data resembling real ones can be simulated by using a detailed model and can then be analyzed with models comprising fewer parameters or even a different kinetic structure. If release rates obtained from the analysis are close to the known ones used for the simulation, it seems justified to substitute the simpler model for the more complex one. In applications this may offer the possibility of using a model in which all parameters are identifiable from the given data.

We demonstrate this approach with two examples shown in Fig. 11. As before, we used the release rates of Fig. 2 Aand generated free calcium transients with the model displayed in Fig. 2 B.

For the model used in the removal fit analysis, we used a different structure. The rate constant k_{ns} was changed to a function that decreased with the amount of released calcium. For this purpose the fractional saturation *f* of the site *S* was calculated (Eq. 13, obtained from Eq. 10 after dividing by S_{total}) and was assumed to modify the uptake rate according to Eq. 14, which replaces Eq. 11:

$$\hat{f} = k_{\text{on}} \cdot \text{Ca}_{\text{free}} \cdot (1 - f) - k_{\text{off}} \cdot f$$
(13)

$$\dot{\mathbf{C}}\mathbf{a}_{up} = (a - f)k_{ns}^0 \cdot \mathbf{C}\mathbf{a}_{free},$$
 (14)

i.e., ak_{ns}^0 is the effective uptake rate constant at zero saturation of *S*, whereas $(a - 1)k_{ns}^0$ is the value at full saturation.

The new model reproduced the slowing of relaxation of the calcium transients, which in the original scheme resulted



FIGURE 11 Effects of misspecified removal models on the estimated release (*A*) Synthetic free calcium transients (Fig. 1 *C*) and removal fit of the misspecified model I (*bold lines*). Best-fit parameters: $k_{ns}^0 = 944.5$, $k_{on} = 10.88$, a = 1.40. (*B*) True release rate (*dashed lines*) (Fig. 1 *A*) and estimation based on the misspecified model I (*solid lines*). (*C*) Synthetic free calcium transients (Fig. 1 *C*) and removal fit of the misspecified model II (*bold lines*). Best-fit parameters: $k_{ns}^0 = 990.5$, $k_{on} = 3.21$. (*D*) True release rate (*dashed lines*) (Fig. 1 *A*) and estimation based on the misspecified model II (*solid lines*).

from the progressive filling of the slow buffer S. In the new scheme, S was eliminated as a buffer by assuming that S_{total} had a negligibly small value. Thus calcium bound to the slow system S does not influence the calcium distribution directly. The uptake in the new scheme can be envisaged to become partially inactivated by a calcium-dependent mechanism that contributes a negligible number of calcium-binding sites. For a further simplification, we set the parameter k_{off} to zero. Thus the resulting model (model I) contained one parameter less than the model used for generating the data.

Fig. 11 *A* displays the calcium transients and the removal fits. Fig. 11 *B* compares the synthetic release rates (*dashed lines*) and those calculated based on the simpler model (*solid lines*). Apart from a slight underestimation of the plateau, the time courses are in good agreement. Fig. 11 *C* shows the removal fits for the case that the parameter *a* in Eq. 14 is fixed to 1 (model II), which means that uptake is completely suppressed when S is fully saturated. The quality of the fit is only slightly worse than that in Fig. 11 *A*, but the estimated release rates in Fig. 11 *D* differ qualitatively from the original ones in showing an increase instead of a steady level and negative going phases at the end of the shorter depolarizations.

DISCUSSION

The determination of the Ca²⁺ release rate from the sarcoplasmic reticulum has been used frequently to characterize excitation-contraction coupling in muscle cells (Baylor et al., 1983; Melzer et al., 1984, 1987; Schneider et al., 1985; Brum et al., 1988b; Sipido and Wier, 1991; Jacquemond and Schneider, 1992; García and Schneider, 1993; Gonzáles and Ríos, 1993; Pape et al., 1993, 1995; Delbono, 1995; Delbono et al., 1995; Delbono and Meissner, 1996; Shirokova et al., 1996; Jong et al., 1996). An empirical determination requires several steps of analysis, including the fit of a removal model to experimental calcium transients (Melzer et al., 1986, 1987; Brum et al., 1988a; Gonzáles and Ríos, 1993; Shirokova et al., 1996).

In the present work we examined, under defined conditions, the degree to which the analysis gives correct results. For this purpose, we applied known release rate waveforms to a known, completely identifiable calcium turnover model, so that the result of the analysis could easily be compared with the true data, and the influence of various disturbing factors on the analysis result could be studied. We described several mathematical tools that can help to improve the method.

The main questions studied were the following: 1) How can the nonlinear removal fit be optimized? 2) To what degree can the release determination cope with observational noise in the calcium records? 3) How sensitive are the results to a violation of the main assumption of the removal fit analysis, namely, that release is completely turned off after repolarization? 4) Is it possible to use simplified removal models comprising fewer parameters?

There is no general procedure that prevents divergence of the removal model fit or convergence to a local minimum of the parameter space. Compared with the classical initial value approach, the multiple shooting algorithm that divides the fit interval into smaller subintervals showed clear advantages. It reduces the inherent numerical instability of the initial value approach, which results from its high sensitivity to the initial choice of the parameter values. The multiple shooting procedure requires less effort to search for a suitable set of initial parameter guesses, because it converges to the correct values for a larger range of initial guesses than the initial value approach. It seems particularly useful when noisy records are to be analyzed. By including the measured concentration values at several points of time during the free calcium decay as initial values for a discontinuous initial trajectory, the multiple shooting fit uses the available dynamic information of the system more efficiently. The reformulation of the initial value problem into a multipoint boundary value problem forces the trajectory to stay close to the measured data, and therefore reduces the chance of divergence, even for ill-chosen starting values. Because the actual linearized problem that has to be solved has the same dimension as the initial value problem, the multiple shooting algorithm does not require significantly more calculation time. We think that this method will be of advantage in applications where short lifetimes of the preparation (e.g., patch-clamp experiments on isolated myocytes or neurons) prohibit time-consuming signal averaging.

The physics underlying the Ca^{2+} compartment model imposes certain constraints on the parameters. Most notably, rate constants and concentrations must be nonnegative. Numerical experience shows that fitting algorithms tend to diverge if these constraints are violated. Therefore, it seems natural to implement them as part of the fitting procedure. Our simulation studies showed that positivity constraints on the parameters do indeed stabilize the fitting procedure substantially.

In the presence of noise, convergence to a local minimum was not a serious problem with the multiple shooting approach, whereas this could easily occur in the initial value approach. With regard to the latter, one additional point should be noted. It has been common practice in previous work with the removal fit method to fix the first point of the fitted trajectory to the observation at the beginning of the fitting interval. However, there is no real justification for this procedure. Instead, the initial value should be treated as an additional free parameter to be fitted. Indeed, our simulations showed that fixing the first point destabilized the procedure, both in the absence and even more in the presence of noise.

Noise is even more of a problem when numerical derivatives have to be calculated. This is required in all available methods of determining the release rate, be they deductive (Baylor et al., 1983) or inductive (Melzer et al., 1987; Brum et al., 1988b). Furthermore, it is necessary when kinetic limitations of indicator dyes have to be corrected for (Klein et al., 1988). There are definitely limits to off-line analysis as a corrective for noise, and experiments should be designed to reduce it as much as possible beforehand. Among the algorithms we compared, the kernel estimator with locally adaptive optimization of the filter bandwidth showed the best results in preserving fast changes and flat sections of the release rate. Difference approximations of the time derivative should only be used when the records have a high signal-to-noise ratio.

The quality of the estimated release rate time course depends not only on the numerical methods applied, but also on the validity of the central assumption that there is no residual release during the decay phase of the calcium signal that is used for the removal model fit.

We investigated the effect of a violation of this assumption by allowing increasing overlap between the analysis interval and slowly decaying release. The calculated release rate underestimated the true one and was temporally distorted. This showed up in an increased ratio of peak to steady level and in a shallow minimum of the release rate right after the peak (Fig. 9 D). From an analysis of the temporal behavior of the model compartments, this effect could be explained by an underestimation of the slow saturable removal component. Overall, however, the main temporal characteristics were rather well preserved, even in cases where the overlap was large.

Larger models including more details of the calcium turnover kinetics than the one used here have been applied to semiempirically derive the rate of release in muscle fibers (Brum et al., 1988b; Gonzáles and Ríos, 1993). In these models, not all of the kinetic parameters can be simultaneously identified, and therefore only a subset of parameters can be obtained by the fit, whereas others have to be fixed beforehand. Moreover, models realistically describing the intracellular situation during Ca release will also have to include diffusion of both Ca and some of its target site populations, and will have to appreciate the fact that the myoplasm is not uniform in terms of Ca release (Ríos and Stern, 1997). Ca is released from distinct regions within the sarcomere, which can create substantial local concentration gradients.

For the purpose of this paper, i.e., for demonstrating the advantages of improved numerical methods, we chose the present, relatively unsophisticated calcium distribution model, simply because of the small number of model parameters involved. However, the general conclusions regarding the benefits of the multiple shooting algorithm combined with an adaptive kernel estimator for deriving the rate of release also hold for more complicated reaction schemes introduced in previous publications.

The problem of actually selecting a minimal kinetic scheme for the removal model fit appropriate for given experimental results was not the scope of this study. However, with the example of Fig. 11, we demonstrate that the approach of analyzing artificial free calcium transients simulated with known release rates may be applied for the purpose of model reduction, i.e., to decide if a given model can be replaced by a simpler one. A simpler model can be trusted to fulfill its task equally well if it reliably returns the known release rates used for the simulation. Conclusions exclusively based on the apparent fit quality, however, may be misleading. The result of Fig. 11, C and D, indicates that a satisfactory fit does not necessarily guarantee perfect reconstruction of the underlying release rate waveform. We therefore feel that one should be cautious in overinterpreting small kinetic details of the release rates determined from muscle cells, and that the general issue of optimal model selection requires further mathematical investigation.

In summary, we showed that advanced methods of data analysis for fitting ODEs as well as for estimating derivatives from noisy data allow for a more reliable reconstruction of the release rate time course than the commonly applied methods. Because the methods investigated here showed their full power in the presence of noise, they will be particularly useful when the time-resolved transmembrane calcium flux is to be determined from calcium transients of smaller cells (cultured myocytes, neurons, etc.) that produce signals with lower signal-to-noise ratios than skeletal muscle fibers.

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